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Calix[4]amido crown functionalized visible sensors for cyanide and iodide anions†

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This study comprises the design and development of calix[4] arene-amido-based ionophores by varying structural stringency and steric hindrance at the lower rim to probe the anion sensing properties. The ionophores are prepared, purified, and characterized using various analytical techniques. The molecular structure of the most active ionophore I is established by single-crystal X-ray characterisation. Out of various anions investigated, iodide and cyanide show the highest sensitivity towards the ionophores investigated. Both anions are sensitive enough to give a visibly distinct color change. The binding properties of the ionophores are established with ^1H & ^{127}I NMR, fluorescence, and UV-vis spectroscopy, revealing that three ionophores strongly interact with CN^- and I^- . The binding constants are calculated via Benesi–Hildebrand plots using absorption data. The time-dependent ^1H NMR revealed strong hydrogen bonding between the OH and NH groups of the ionophore and cyanide anion. The ^{127}I NMR shows the highest 27.6 ppm shift after 6 h for ionophore I. The crystal structure revealed hydrogen bonding of N–H protons of the amide pendulum and phenolic oxygen of the calix rim. The Job's plot depicted the possibility of a 1 : 1 complex of ionophores with both anions.

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Introduction

Supramolecular based anion-selective visual sensors have drawn significant attention in the past two decades¹ due to their vital role in the environment and biological systems. The detection of anions is more challenging than cations due to their bigger size, structural specificity, basicity, and nucleophilicity². Among various water-soluble anions, biologically important cyanide (CN^-) and iodide (I^-) are of great interest. Cyanide is highly toxic because of its deleterious effects on human health in minute quantities. Cyanide is also involved in chemicals, polymers, metal mining industries, and chemical warfare agents.³ As per the WHO guidelines, the maximum allowed cyanide concentration in drinking water is two ppb per liter.^{4,5}

In contrast, iodine is an essential micronutrient for the human body, responsible for producing thyroid hormones. Iodine deficiency can also cause enlarged thyroid glands, thyroid cancer, and pregnancy complications for babies. Excessive ingestion can lead to hyperthyroidism diseases like Graves' disease.⁶ Thus, there is a dire need to develop easy, quick, and selective sensors in diverse samples.⁷ Despite the

fact that several instrumental methods have been developed for the detection of cyanide and iodide ions, they are time-consuming, expensive, and cumbersome due to portability, calibration, and sample preparation. Hence there is a great need to develop colorimetric chemosensors for anions like cyanide and iodide, which are low cost and nontoxic and can be used with ease. Recently impressive advances have been made towards the development of visible cyanide sensors introducing molecules and materials such as pillararene,⁸ naphthofuran carbohydrazide,⁹ supramolecular gel,¹⁰ naphthalene,¹¹ cyclodextrin,¹² curcumin¹³ chromone,¹⁴ pyrene-thiacalix[4]arenes,¹⁵ calix[4]arene–naphthalimide,¹⁶ calix[3]pyrrole,¹⁷ Schiff bases,¹⁸ nitrobenzoxadiazole,¹⁹ and organogelators.²⁰ For iodide detection, gold nanoparticles,²¹ poly(vinylpyrrolidone)-supported copper nanoclusters,²² bidentate ureido-dihomooxacalix[4]arene,²³ hexahomotrioxacalix[3]arene,²⁴ and porous ionic polymer⁷ based colorimetric sensors are noteworthy. Recently, we have reported dipicryl hydrazine based colorimetric sensors for selective determination of anions like fluoride, acetate, hydroxide, cyanide, and hydrogen sulfate. The anion binding ability of dipicrylhydrazine with various anions like fluoride, acetate, hydroxide, cyanide, and hydrogen sulfate is due to polar non-polar interactions and hydrogen bonding.²⁵ Continuing our efforts, hereby we are reporting furfuryl and benzyl functionalized calix[4]amido crown-based visual molecular sensors for cyanide and iodide ions with ppm level detection limit. Limited studies pertain to the application of calix[4]amido crowns have been explored for the detection of transition metal

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cations²⁶ and limited anions.²⁷ The amide moiety of calix[4]amido crowns have a unique role in sensory behavior since N–H can interact with anions *via* selective hydrogen bonding donor.^{28–31} The furfuryl and benzyl substituent attach with amide moiety provide secondary interactions with chromophoric assistance, essential for the colorimetric sensor. These studies are unexplored in the context of cyanide and iodide. The current study also throws the light on detailed synthesis of furfuryl and benzyl functionalized calix[4]amido crowns, their stereochemistry, rigidity, steric hindrance, and interaction behavior using diverse analytical techniques such as NMR, XRD, and UV-vis.

Experimental section

Experimental materials

All reactions and manipulations were routinely performed under an inert atmosphere. Solvents such as toluene, ethanol, tetrahydrofuran, acetonitrile were purchased from Merck and purified by standard procedures, and freshly distilled before use. Reagents such as *p*-*tert*-butyl phenol and diphenyl ether, ethyl bromoacetate, chloroacetylchloride, and diethylenetriamine were purchased from Sigma-Aldrich and used as received. The furfuryl amine, benzylamine, and all tetrabutylammonium salts of anions (F[−], Cl[−], Br[−], I[−], CN[−], OH[−], CH₃COO[−], NO₃[−], ClO₄[−] and HSO₄[−]) were purchased from Alfa-Aesar. Analytical thin-layer chromatography was carried out on silica plates (SiO₂, Merck 60 F₂₅₄) Obtained from E. Merck Chemical Co.

Physical measurement

UV-vis spectra were recorded using a Varian model Cary Win 400 UV/Vis spectrophotometer. The Perkin Elmer LS55 fluorescence spectrophotometer was used for recording fluorescence spectra. Nuclear magnetic resonance spectra (¹H NMR & ¹³C NMR) were recorded on a Bruker 500 MHz WB FT-NMR spectrometer having proton noise decoupling mode with a standard 5 mm probe. Chemical shifts for ¹H NMR spectra are reported as δ in parts per million (ppm) downfield from SiMe₄ (δ 0.0) and relative to the signal of chloroform-d (δ 7.26, singlet). Multiplicities were given as: s (singlet); d (doublet); t (triplet); q (quartet) or m (multiplets). For proving strong binding between ionophore hosts and anionic guests, ¹²⁷I NMR was recorded separately at 100 MHz under same solvent mixture ratio. The melting point was checked by Buchi melting point apparatus M-565. Mass spectra were recorded by Bruker microTOF-Q II using ESI source, and elemental analysis was done on FLASH EA 1112 series from Thermo finnigan, Italy.

Synthesis and characterization of ionophores

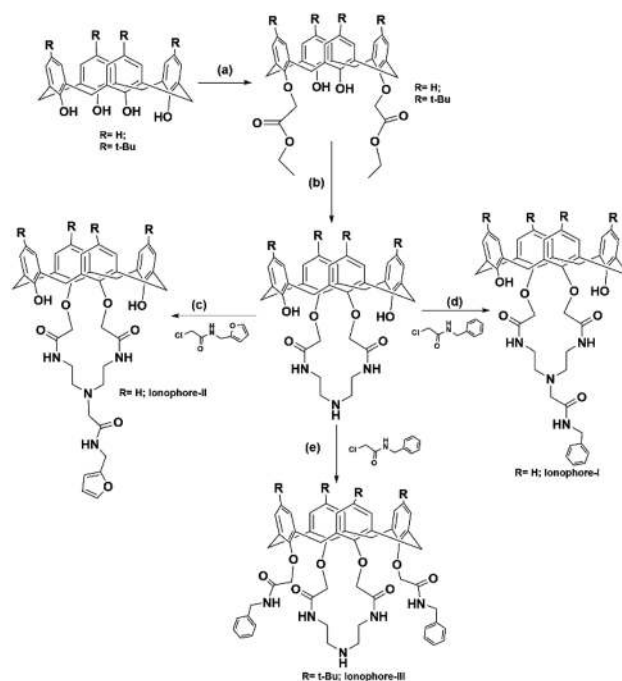
The *p*-*tert*-butylcalix[4]aene, calix[4]arene, 1,3 diester calix[4]arene and calix[4]amido crown-5 were prepared as per the reported literature procedure.^{26a,32–34} *N*-Furfurylchloroacetamide and *N*-benzylchloroacetamide reagents were synthesized according to reported methods.^{35,36}

Synthesis of ionophore I

To a solution of calix[4]amidocrown-5 (0.91 g, 1.5 mmol) in dry CH₃CN (70 mL) were added *N*-benzyl chloroacetamide (0.83 g, 4.5 mmol) and K₂CO₃ (1.04 g, 7.5 mmol). The reaction mixture was refluxed for 48 h. The reaction mixture was quenched by addition of 5% HCl (10 mL) and CH₂Cl₂ (40 mL). The organic phase was separated and washed with water (3 × 40 mL), dried over anhydrous sodium sulphate, and evaporated *in vacuo*. The crude product was recrystallized with CHCl₃ and methanol to yield a white crystalline compound (0.89 g, 78.95%) as shown in Scheme 1. mp 229 °C, ¹H-NMR: (500 MHz, CDCl₃, see ESI Fig. S1†) δ 8.82 (t, 2H, CONH), 7.80 (s, 2H, OH), 7.64 (t, 1H, NH), 7.37–7.31 (m, 5H, benzyl), 7.13 (d, 4H, ArH), 6.95 (d, 4H, ArH), 6.85 (t, 2H, ArH), 6.78 (t, 2H, ArH), 4.48 (d, 2H, benzyl), 4.40 (s, 4H, OCH₂), 4.08 (d, 4H, ArCH₂Ar AB system), 3.53 (br s, 4H, CONHCH₂), 3.48 (d, ArCH₂Ar, AB system), 3.26 (s, 2H, benzyl), 2.93 (br, s, 4H, CONHCH₂CH₂). ¹³CNMR: (see ESI Fig. S2†); δ 71.1, 168.7, 151.9, 150.9, 132.5, 129.7, 129.2, 129.1, 128.8, 128.6, 128.5, 127.9, 127.7, 127.6, 127.2, 126.9, 126.7, 120.9 (ArCH, ArC, CONH), 74.8 (OCH₂), 59.9, 54.9 (benzylC), 44.19, 43.2, 39.9, 37.1, 31.9, 31.5 (CONHCH₂CH₂, ArCH₂Ar), 30.3, 30.1, 29.7, 29.4, 27.1, 22.7, 19.8, 14.1 (benzylC). Maldi TOF MS ES+ (*m/z*) calculated for C₄₅H₄₆O₇N₄, 754.34, found 755.34 [M + H]⁺, 756.34 [M + 2H]⁺ (see ESI Fig. S14†) CHN Anal. calcd for C₄₅H₄₆O₇N₄: C 71.60, H 6.14, N 7.42, O 14.84; found: C 71.59, H 6.12, N 7.41, O 14.82.

Synthesis of ionophore II

To a solution of calix[4]amidocrown-5 (0.91 g, 1.5 mmol) in dry CH₃CN (70 mL) were added *N*-furfurylchloroacetamide (0.78 g,



Scheme 1 Showing synthetic procedure for compounds: (a) K₂CO₃, BrCH₂COOC₂H₅, acetone; (b) toluene/ethanol, diethylenetriamine; (c and d) K₂CO₃, acetonitrile, 48 h, reflux (e) NaH, THF: DMF: 2.5 : 1, 72 h reflux.





Fig. 1 Colorimetric analysis for (a) ionophore I, (b) ionophore II, and (c) ionophore III upon addition of different anions.

4.5 mmol) and K_2CO_3 (1.04 g, 7.5 mmol). The reaction mixture was refluxed for 48 h. The solvent was removed *in vacuo* and the residue was quenched by addition of 5% HCl (10 mL) and CH_2Cl_2 (40 mL). The organic phase was separated and washed with water (3×40 mL), dried over anhydrous sodium sulphate, and evaporated *in vacuo*. The crude product recrystallized with $CHCl_3$ and methanol to yield a white crystalline compound (0.80 g, 72.8%) as shown in Scheme 1. mp 258 °C; 1H NMR: (500 MHz, $CDCl_3$ see ESI Fig. S3†); δ 8.76 (t, 2H, CONH), 7.79 (s, 2H, OH), 7.54 (t, 1H, NH), 7.12 (m, 1H, furfuryl) 7.07–7.05 (d, 4H, ArH), 6.90–6.88 (d, 4H, ArH), 6.78 (t, 2H, ArH), 6.71 (t, 2H, ArH), 6.18 (s, 2H, furfuryl), 4.45 (s, 4H, OCH_2), 4.39 (d, 2H, CH_2 furfuryl), 4.09 (d, 4H, $ArCH_2Ar$, AB system), 3.47 (d, 4H, $ArCH_2Ar$, AB system), 3.42 (d, 4H, CH_2), 3.18 (br s, 2H, CONHCH₂), 2.82 (brs, 4H, CONHCH₂CH₂). ^{13}C NMR: (see ESI Fig. S4†): δ 171.1, 168.7, 152.2, 150.9, 141.9, 132.5, 129.7, 129.2, 127.6, 126.7, 120.9 (ArCH, ArC, CONH), 110.5, 106.8 (furfuryl C), 74.9 (OCH_2), 59.61, 54.92 (furfuryl C), 39.9, 36.3, 31.9, 31.5 (CONHCH₂CH₂, $ArCH_2Ar$), 29.7, 22.70, 14.1 (furfuryl C). Maldi TOF MS ES+ (m/z) calculated for $C_{43}H_{44}O_8N_4$, 744.32, found 745.32 [$M + H^+$]⁺, 746.32 [$M + 2H^+$]⁺, 747.32 [$M + 3H^+$]⁺ (see ESI Fig. S15†), CHN Anal. calcd for $C_{43}H_{44}O_8N_4$: C 69.34, H 5.95, N 7.52, O 17.18; found: C 69.30, H 5.91 N 7.41, O 17.10.

Synthesis of ionophore III

To a solution of *p*-tert-butyl calix[4]amido crown-5 (1.24 g, 1.5 mmol) in dry THF (50 mL) and dry DMF (20 mL) under N_2 was added NaH (0.18 g, 7.5 mmol). The mixture was stirred at RT for 30 min followed by addition of *N*-benzyl chloroacetamide (0.83 g, 4.5 mmol). The reaction mixture was refluxed for 72 h, the solvent was removed *in vacuo* and quenched by the addition of 5% HCl (10 mL) and CH_2Cl_2 (40 mL). The organic phase was separated and washed with water (3×40 mL), dried over anhydrous sodium sulfate and evaporated *in vacuo*. The crude product recrystallized with $CHCl_3$ and methanol to yield a white crystalline compound (1.126 g, 67.42%) as shown in Scheme 1.

mp 244 °C; 1H -NMR: (500 MHz, $CDCl_3$ see ESI Fig. S5†); δ 8.82 (t, 2H, CONH), 7.66 (t, 1H, CONH), 7.41 (s, 2H, ArH), 7.28 (m, 6H, benzyl), 7.20 (m, 4H, ArH), 7.11 (s, 4H, ArH), 6.85 (s, 4H, ArH), 4.47 (d, 2H, benzyl), 4.38 (s, 4H, OCH_2), 4.04 (d, 4H, $ArCH_2Ar$ AB system), 3.51 (br s, 4H, CONHCH₂), 3.45 (s, 4H, CONHCH₂), 3.40–3.37 (d, $ArCH_2Ar$, AB system), 3.26 (s, 2H, benzyl), 2.91 (br, s, 4H, CONHCH₂CH₂), 1.63 (1H, NH), 1.30 (s, 18H, *t*-butyl), 0.99 (s, 18H, *t*-butyl). ^{13}C NMR: (see ESI Fig. S6†); δ 170.1, 167.9, 148.4, 147.6, 147.5, 142.53, 137.86, 130.9, 127.5, 126.5, 126.4, 126.1, 125.1, 124.6 (ArCH, ArC, CONH), 73.6 (OCH_2), 58.7, 53.9 (benzylC), 42.16, 38.5, 37.1, 33.1, 32.9, (CONHCH₂CH₂, $ArCH_2Ar$), 30.7, 30.5, 29.8, 29.7, 28.6, 21.8, 13.1 (benzyl C). Maldi TOF MS ES+ (m/z) calculated for $C_{70}H_{87}O_8N_5$, 1125.66, found 1126.66 [$M + H^+$]⁺, 1127.66 [$M + 2H^+$]⁺ (see ESI Fig. S16†), CHN Anal. calcd for $C_{70}H_{87}O_8N_5$: C 74.6, H 7.78, N 6.22, O 11.36; found: C 74.9, H 7.12, N 6.41, O 11.82.

Visual detection experiment

Visual detection experiments were carried out for recognizing the host–guest interactions of ionophores I–III with various anions. In a typical experiment, a series of stock solutions of anions (5×10^{-3} M) and ionophores (5×10^{-5} M) were prepared in freshly distilled acetonitrile. Further, 2 mL aliquots of stock solution were taken and mixed with 2 mL of the anion solution. Typical photographs were taken with a digital camera. All ionophores gave instant color change for iodide and cyanide ions. Iodide and cyanide ion gave pink and wine color for ionophore I, respectively, consistent with the color variation displayed in Fig. 1. The ionophore II showed light pink color with iodide while cyanide turned yellow. In contrast to ionophore I and II, ionophore III gives light yellow color with cyanide, but there is no visible change in the case of iodide.

Interaction with anions by UV-vis study

In UV-vis titration experiment, 1 mL (5×10^{-3} M) stock solution of tetrabutylammonium anions (F^- , Cl^- , Br^- , I^- , CN^- , OH^- , CH_3COO^- , NO_3^- , ClO_4^- and HSO_4^-) was added to a 1 mL (5×10^{-5} M) acetonitrile solution of ionophore I–III in ascending order in ten fractions (1 : 100 ratio). The spectral changes were recorded to ascertain the interaction/selectivity. Based on spectral changes, titrations were performed with strongly interacted anions cyanide and iodide to investigate the binding sites and determine the binding constant. We have performed competitive binding with other anions (F^- , Cl^- , Br^- , I^- , CN^- , OH^- , CH_3COO^- , NO_3^- , ClO_4^- and HSO_4^-) and found that ionophores are showing selective binding and color change towards cyanide.

NMR spectral study

The 1H and ^{13}C NMR spectra of ionophore I–III were recorded before and after the addition of selected anions (CN^- and I^-) in $CDCl_3$: CD_3CN (1 : 2.5). In this experiment, 0.003 mmol of ionophore I–III were dissolved separately in 0.5 mL mixture containing $CDCl_3$: CD_3CN (1 : 2.5), and further, add various anions (20 molar equivalents) in the form of TBA salts. Proton



NMR was recorded at different time intervals up to 24 h after the addition of anions.

Structure determination by single-crystal X-ray study

Appropriate crystal was selected for ionophore **I**, dipped in paratone oil, and mounted on cryo loop. Crystal data were collected at 100 K using graphite monochromatic MoK α ($\lambda = 0.71073$ Å) radiation on a Bruker SMART APEX diffractometer equipped CCD area detector. The SAINT software³⁷ was used for data integration and reduction. Empirical absorption correction was applied to the collected reflections with SADABS.³⁸ SHELXTL³⁹ was used to solve the structures by direct methods using and refined on F_2 by the full-matrix least-squares technique using the SHELXL-97 (ref. 40) package. Non-hydrogen atoms were refined anisotropically till convergence is reached. The MERCURY 3.8 (ref. 41) is used to generate graphics. The X-ray structure of ionophore **I** is shown in Fig. S17,[†] wherein chloroform and water molecules have been omitted for clarity. The phenolic OH of the alternate phenyl rings of the calix moiety are substituted by amide-linked chains. The presence of hydrogen bonding between amide NHs and phenolic oxygen atoms is also observed. Crystallographic parameters for the ionophore **I** are given in Table S1.[†]

Result and discussion

UV-vis absorption studies

Quantitative investigations were carried out to understand cyanide and iodide anions' encapsulation behavior, in presence of different ionophores using a spectrophotometric titration method in CH₃CN. The absorption spectra of ionophores **I**, **II**, and **III** at variable concentrations of iodide and cyanide are shown in Fig. 2 and 3. Ionophores **I** and **II** show a strong absorption maximum at 279 nm, while **III** shows at 285 nm. Single spectral band for anions and dual spectral bands were observed for all ionophores from 273 to 290 nm. Upon increasing the anions' concentration, the absorbance of ionophores is increased at both the maxima as per Fig. 2a–c (for iodide) and Fig. 3a–c (for cyanide). In the absence of ionophore, iodide and cyanide anions show their respective peaks at 249 and 275 nm, respectively. The absorbance increased with the concentration of anions due to interaction with ionophores. Changes in the absorbance of ionophores (measured against the solvent as reference) upon the addition of anions were observed at 279 nm for ionophore's **I** and **II**, and 285 nm for ionophore **III**. Owing to their hydrophobic cavities, host ionophores offer the iodide and cyanide a suitable environment for

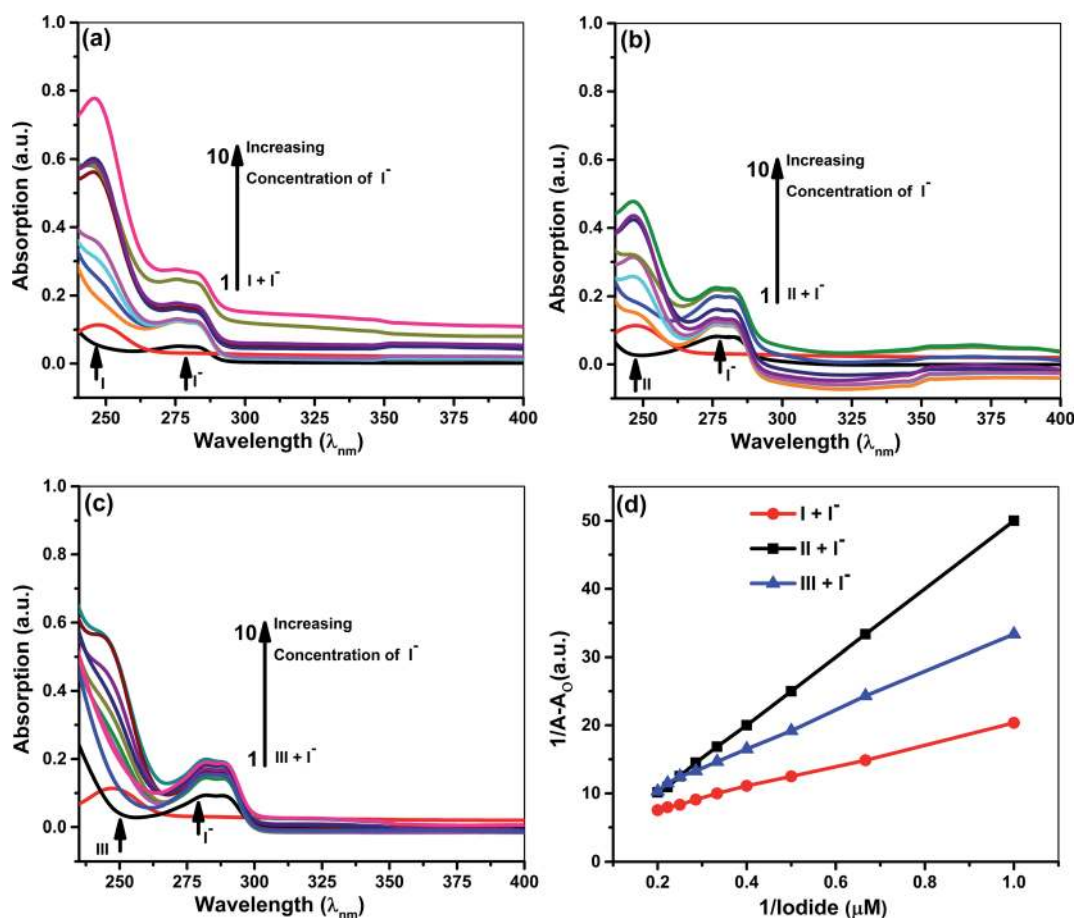


Fig. 2 Absorption spectra of 5×10^{-5} M ionophores in the absence and presence of iodide at concentrations 5×10^{-5} to 5×10^{-6} M (a) ionophore I (b) ionophore II (c) ionophore III (d) Benesi–Hildebrand absorption plot for 1 : 1 complexation of iodide with ionophores I–III.



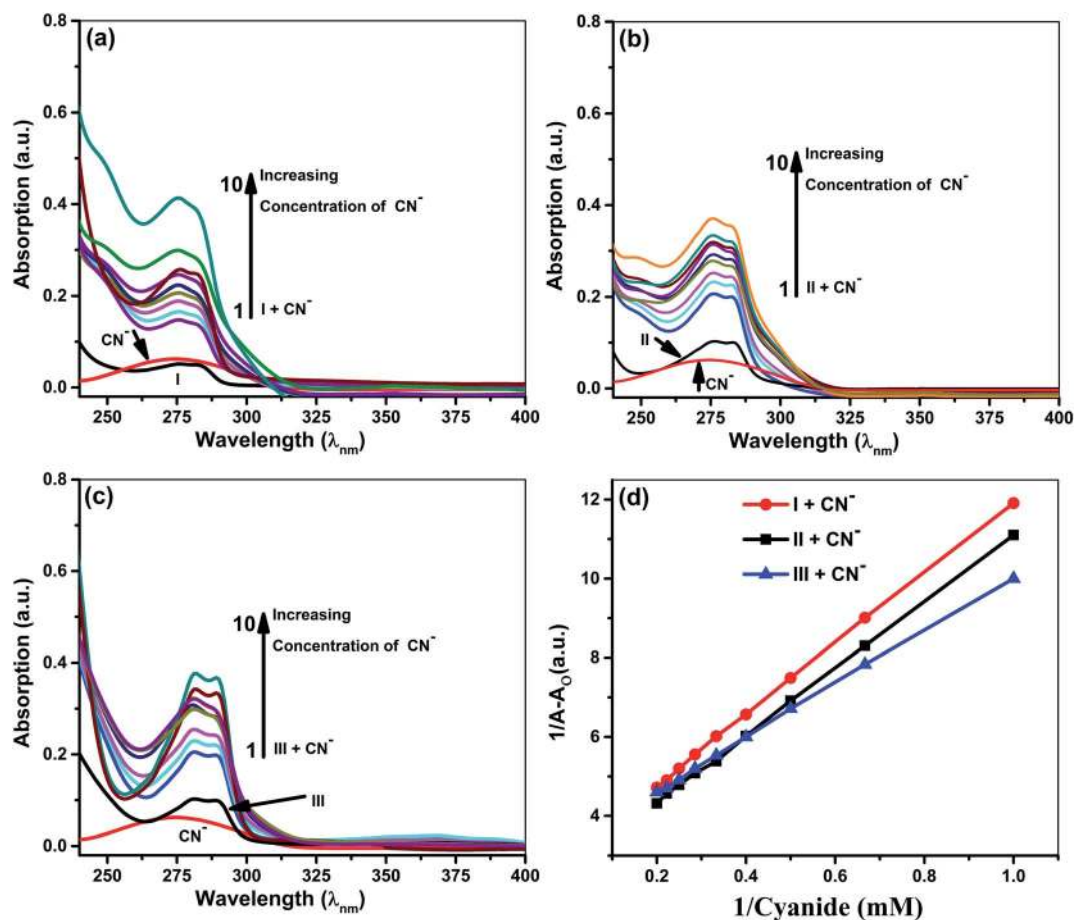


Fig. 3 Absorption spectra of 5×10^{-5} M ionophore's in the absence and presence of cyanide at concentrations 5×10^{-3} to 5×10^{-4} M (a) ionophore I (b) ionophore II (c) Ionophore III (d) Benesi–Hildebrand absorption plot for 1 : 1 complexation of cyanide with ionophores I–III.

interaction to form inclusion complexes. The binding constants for the formation of ionophores: iodide and ionophores: cyanide complexes are determined by analyzing the absorbance changes. The binding constant K_a and stoichiometry of the inclusion complexes of ionophores with iodide and cyanide can be determined by the Benesi–Hildebrand equation (Table 1).⁴² For ionophores I, II and III, the calculated binding constant from the straight-line slope for iodide is found to be 552 M^{-1} , 509 M^{-1} , and 350 M^{-1} , respectively. At the same time, for

cyanide, the values are lower, *i.e.*, 319 M^{-1} , 304 M^{-1} and 209 M^{-1} , respectively, at 298 K. This behavior explains strong complexation towards both anions in the following order: **I** > **II** > **III**. It is observed that ionophore I gave a stronger K_a value than II and III, indicating its superior interaction compared to heavily alkylated and arylated II and III. Considering the structural features of the host and guest, we presumed that the deprotonation and charge transfer *via* polarization leads to complexation with iodide. In contrast, hydrogen bonding

Table 1 Binding constants of the complexes *via* absorption spectral maxima for iodide and cyanide anions with Ionophores I, II and III

S. No.	Ionophores	λ (nm)	Monitored λ_{max} (nm)	Binding constant K_a (M^{-1})	R^2	Complex
For iodide anion						
1	I	273–284	279	552.0	0.9988	1 : 1
2	II	273–284	279	509.0	0.9999	1 : 1
3	III	280–290	285	350.0	0.9982	1 : 1
For cyanide anion						
4	I	273–284	279	319.0	0.9996	1 : 1
5	II	273–284	279	304.0	0.9996	1 : 1
6	III	280–290	285	209.0	0.9993	1 : 1



Table 2 Binding constants of the complexes *via* emission spectral maxima for iodide and cyanide anions with ionophores I, II and III

S. No.	Ionophores	λ (nm)	Monitored λ_{\max} (nm)	Binding constant K_a (M^{-1})	R^2	Complex
For iodide anion						
1	I	375–385	379	834	0.9997	1 : 1
2	II	375–385	379	729	0.9995	1 : 1
3	III	380–384	380	498	0.9990	1 : 1
For cyanide anion						
4	I	375–385	379	956	0.9994	1 : 1
5	II	375–385	379	753	0.9998	1 : 1
6	III	380–384	380	604	0.9991	1 : 1

between cyanide and amines of ionophore leads to deprotonation and strengthens the host–guest association. All the above phenomena are common in all ionophores. Since steric hindrances are in the following order: **III** > **II** > **I**, bind in reverse order.

The plot's good linearity with better correlation co-efficient R^2 more than 0.998 for all complexes shows the strong formation of 1 : 1 complexes between anions and ionophores **I**, **II**, and **III**. Stoichiometric ratios are calculated from Job's Plot, which indicates towards 1 : 1 complex formation (see ESI Fig. S7†).⁴³

Fluorescence studies

Samples prepared for UV-vis analysis were further used for the spectrofluorometric study. Table 2 and Fig. 4 & 5 show the supramolecular interaction of ionophores with iodide and cyanide anions. An enrichment of fluorescence intensity of ionophores was observed upon adding iodide and cyanide aliquots. Such qualitative assessment of the inclusion complexation behavior by spectral titrations shows a single emission maximum at 379 nm for ionophore **I** and **II** and 380 nm for ionophore **III**, respectively, with very low fluorescence

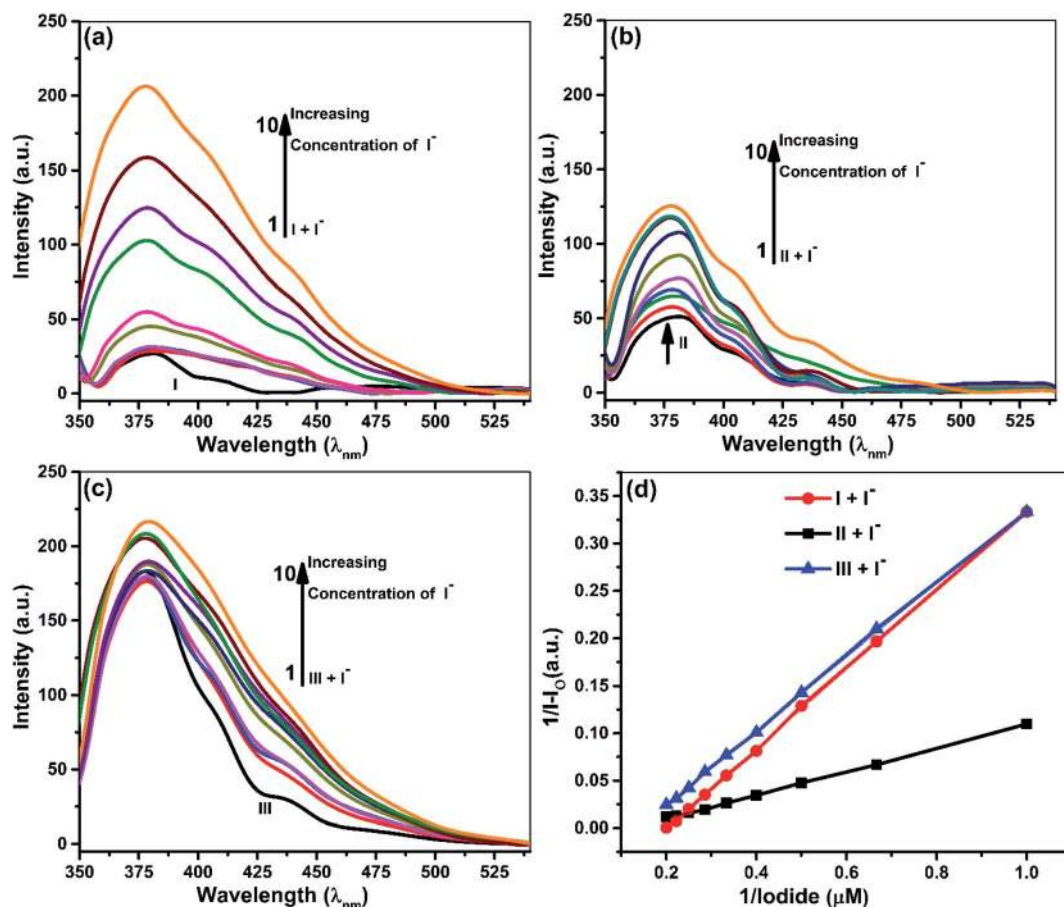


Fig. 4 Fluorescence spectra of 5×10^{-5} M ionophores in the absence and presence of iodide at concentrations 5×10^{-5} M to 5×10^{-6} M. (a) ionophore I (b) ionophore II (c) ionophore III (d) Benesi–Hildebrand emission plot for 1 : 1 complexation of iodide with ionophores I–III.



intensity before the addition of anions at excitation wavelength of 279 nm for **I** and **II** and 285 nm for ionophore **III**. In the case of iodide anion, there is no spectral shift observed in ionophore **II** and **III**, while ionophore **I** shows the blue shift to 375 nm (~ 5 nm). Fig. 4 and 5 shows the emission spectra of iodide and cyanide anion with all ionophores.

Enhancement of emission maxima shows gradual redshift for **I** and **II** from 379 nm to 375 nm, while no noticeable changes were observed for ionophore **III**. Alteration in the host molecule's photophysical and photochemical properties establishes transference of the anions from a more protic atmosphere to a less protic atmosphere, *i.e.*, the cavity of ionophores. The ionophores can form intra and inter-molecular hydrogen bonds between the phenolic O–H and the nitrogen of the cyanide. The supramolecular cavity of ionophores offers a protective environment with excitation of singlet species and vibrational restriction to iodide and cyanide molecules during encapsulation. The study clearly explained that the encapsulation or inclusion phenomenon depends on the fitting and selective detection concept between host and guest molecules. Such encapsulations cannot be clearly explained on single parameter, various spectacles leads to several weak intermolecular forces such as ion–dipole, dipole–dipole, van der Waals, electrostatic

forces, hydrogen bonding strengths, and steric effects. Not only this, hydrogen-bonding donor binding to an anions involves deprotonation of hydrogen binding donor, in which proton transferred to basic anions and it varies with size and electro-negative character of anions CN^- and I^- .

All the above concepts collaborate and contribute to the formation of a stable inclusion. Hence, inclusion formation constants or binding constants (K_a) of all the complexes were calculated from fluorescence data using the Benesi–Hildebrand equation, and the stoichiometric ratio was determined (Table 2).^{42,43} A good linearity with a better regression co-efficient, R^2 is obtained. The binding constant ' K_a ' is calculated from the straight line's slope, considering both ground and excited state measurements. It was found to be 834 M^{-1} , 729 M^{-1} & 498 M^{-1} in the case of iodide anion, while the derived binding constant from cyanide complexes were 956 M^{-1} , 753 M^{-1} & 604 M^{-1} for ionophore **I**, **II** & **III**, respectively. The above behavior indicates that all ionophore forms different inclusion complexes with cyanide and iodide ions. The linearity of the plot shows that the stoichiometry of the complex between ionophores and both the anions was found to be 1 : 1. The binding constant values for ionophore **I** with all anions are higher than other complexes (Table 2), indicating that ionophore's capability to form

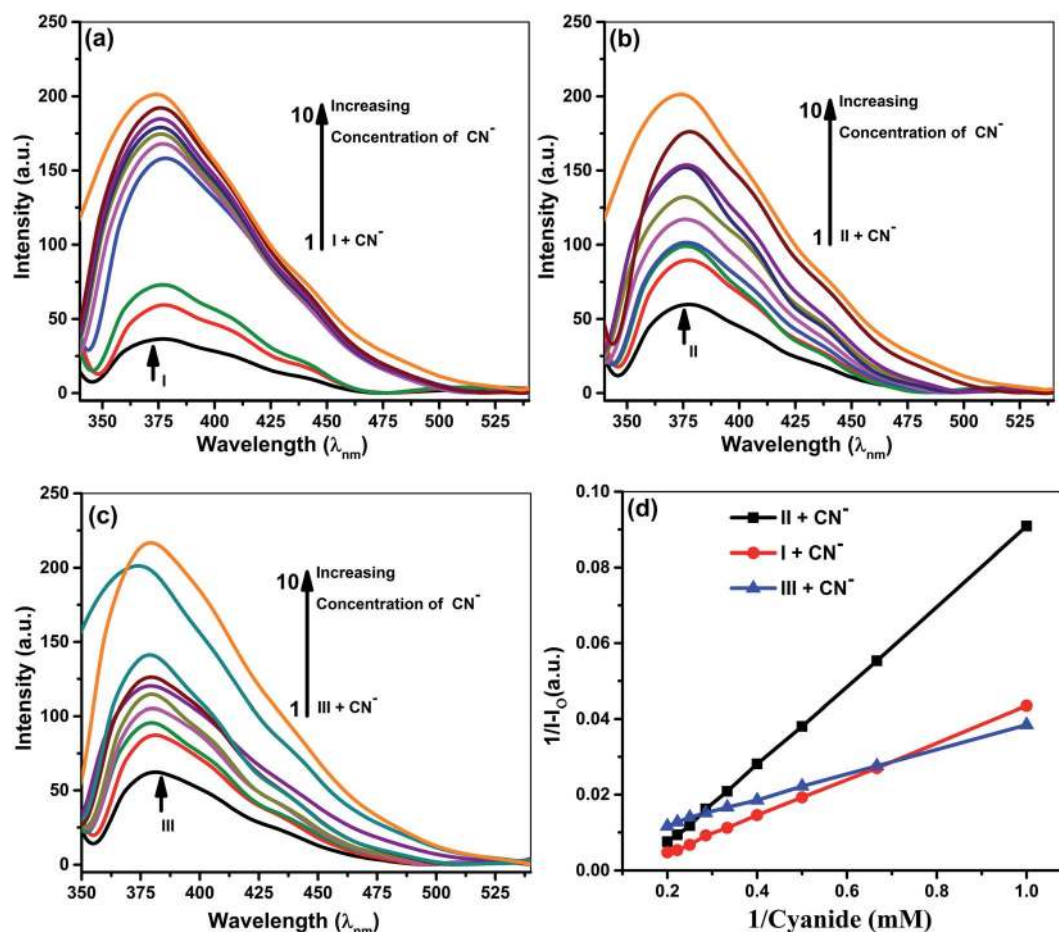


Fig. 5 Fluorescence spectra of 5×10^{-5} M ionophores in the absence and presence of cyanide at concentrations 5×10^{-3} M to 5×10^{-4} M. (a) ionophore I (b) ionophore II (c) ionophore III (d) Benesi–Hildebrand absorption plot for 1 : 1 complexation of cyanide with ionophores.



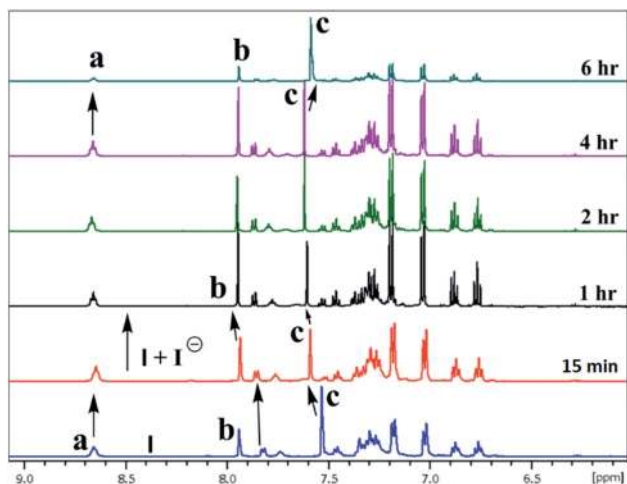


Fig. 6 A part of time-dependent ^1H NMR spectra (500 MHz) in CDCl_3 : CD_3CN at 25°C showing protons of ionophore **I** in the absence presence of iodide anion.

inclusion complexes with iodide and cyanide anions. The ionophore **I** can complex better with the cyanide and iodide than arylated **II** and **III**. For iodide, deprotonation and charge transfer *via* polarization are major interacting forces. However, hydrogen bonding between nitrogen atoms of cyanide and amines of ionophore leads to complete deprotonation and strengthening host-guest association. All above phenomena were common in all ionophores; however, steric hindrances are highest in **III**, which have the least binding. Stoichiometric ratios were calculated from Job's plot that shows 1 : 1 complex formation (see ESI Fig. S7[†]).⁴³ Binding constant data's for emission and absorption data's were different, but dual binding constants and better R^2 values in same sequence clearly explain that ionophores have a strong binding capability with anions.

^1H & ^{127}I NMR study

The inclusion of guest ions inside the ionophore's cavity was ascertained by changes observed in ^1H & ^{127}I NMR. The NMR

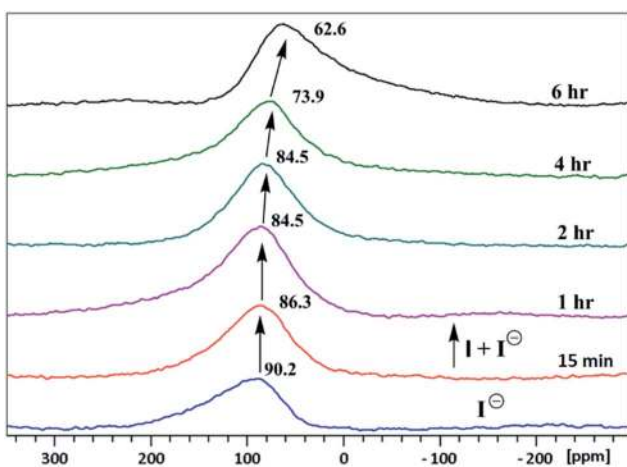


Fig. 7 Time-dependent ^{127}I NMR spectra (500 MHz) in CDCl_3 : CD_3CN at 25°C of tetrabutylammonium iodide with ionophore **I**.

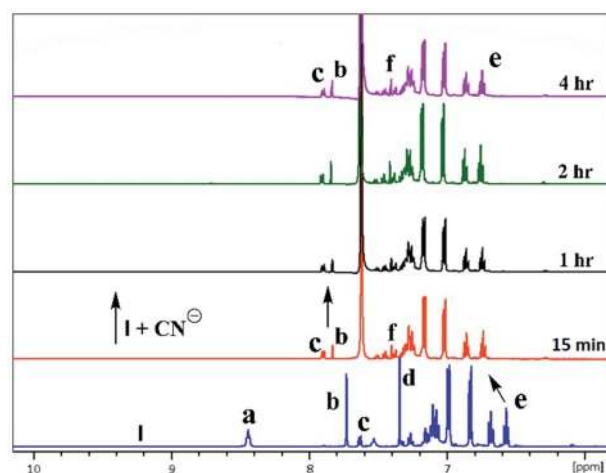


Fig. 8 A part of time-dependent ^1H NMR spectra (500 MHz) in CDCl_3 : CD_3CN at 25°C for tetrabutylammonium cyanide anion with ionophore **I**.

interaction studies were carried out by adding the equimolar concentration of ionophore **I** and anions at 297 K. For iodide anion there is slight significant change was seen in time-dependent ^1H NMR (Fig. 6) due to the large size and poor hydrogen bonding tendency. Peak a get diminished with slight shift in case of peak c and becomes constant after 6 h. Interestingly, the ^{127}I showed 27.6 ppm upfield shift due to inclusion and non-covalent interaction inside the cavity (Fig. 7). No further changes were observed after 6 h. The other two ionophores **II** and **III**, also showed similar trends (see ESI Fig. S8–S13[†]).

In the case of cyanide, clear changes were observed in time-dependent proton NMR with deprotonation of peak 8.45 ppm ($-\text{NH}$) and 7.75 ppm ($-\text{OH}$) of ionophores within 15 min, as shown in Fig. 8. For ionophore **I**, aromatic peaks shifted downfield due to the strong nucleophilicity of cyanide ion inclusion. It has been observed that ionophores do not behave chemidosmetrically as the color started fading after 72 h and the N–H at 8.45 ppm proton started reappearing (Fig. S18[†]). The sterically crowded ionophore **III** showed the least binding capability as observed in ^{127}I NMR shift (see ESI Fig. S13[†]).^{7b}

Conclusions

Three new functionalized furfuryl and benzyl derivated of calix [4]amido crown based molecular ionophores were synthesized to investigate their anion sensing properties. These ionophores were designed with variation in lower rim conformation based on rigidity and steric hindrance at the binding site. A number of anions were investigated with three ionophores indicating that ionophore **I** bind strongly with both iodide and cyanide ions with interaction order $\text{I} > \text{II} > \text{III}$. The ionophore **I** shows drastic color change from colorless to pink and wine red for iodide and cyanide anion. For iodide ion, ionophore **II** and **III** do not show very sharp color change while CN^- gave colorless to light yellow. The strong binding for iodide ion was evident by ^{127}I NMR



where ionophore **I** gave 27.6 ppm upfield shift while for ionophore **II** and **III**, 13.9 and 10 ppm, respectively. The cyanide ion shows strong interaction with N–H and O–H groups, with all three ionophores, resulted in a color change. The color change is attributed to the charge transfer process among the anions and lower amide pendulum. The ionophore **I** interact strongly with iodide and cyanide because of the less hindered binding site, for effective interaction with anions. The binding isotherms were fitted to a 1 : 1 binding model with better binding constants, as suggested by the Benesi–Hildebrand plot for all ionophores.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- I. A. Rather, S. A. Wagay and R. Ali, Emergence of anion-p interactions: The land of opportunity in supramolecular chemistry and beyond, *Coord. Chem. Rev.*, 2020, **415**, 213327.
- R. Ali, R. C. Gupta, S. K. Dwivedi and A. Misra, Excited state proton transfer (ESIPT) based molecular probe to sense F[−] and CN[−] anions through a fluorescence “turn-on” response, *New J. Chem.*, 2018, **42**, 11746–11754.
- (a) C. Young, L. Tidwell and C. Anderson, *Cyanide: Social Industrial and Economic Aspects*, Minerals, Metals, and Materials Society, Warrendale, 2001; (b) Z. Xu, X. Chen, H. N. Kim and J. Yoon, Sensors for the optical detection of cyanide ion, *Chem. Soc. Rev.*, 2010, **39**, 127–137.
- Guidelines for Drinking-Water Quality*, World Health Organization, 20 Geneva, 1996.
- F. P. Simeonova, L. Fishbein, Concise International Chemical Assessment Document 61, *Hydrogen cyanide and cyanides: human health aspects*, World Health Organization, Geneva, 2004, <https://apps.who.int/iris/handle/10665/42942>.
- (a) H. ling and D. Yue, *BMC Pediatr.*, 2020, **20**(169), 1–4, report on Flashimato and Graves; (b) P. Saxena, G. C. El Hamri, M. Folcher, H. Zulewski and M. Fussenegger, Synthetic gene network restoring endogenous pituitary–thyroid feedback control in experimental Graves’ disease, *Proc. Natl. Acad. Sci. U. S. A.*, 2016, **113**, 1244–1249.
- (a) Z. Chen, R. Sun, S. Feng, D. Wang and H. Liu, Porosity-Induced Selective Sensing of Iodide in Aqueous Solution by a Fluorescent Imidazolium-Based Ionic Porous Framework, *ACS Appl. Mater. Interfaces*, 2020, **12**, 11104–11114; (b) P. R. Sharma, S. Pandey, V. K. Soni, G. Choudhary and R. K. Sharma, Macroscopic recognition of iodide by polymer appended calix[4]amidocrown resin, *Supramol. Chem.*, 2019, **31**, 634–644.
- (a) Q. Lin, K. P. Zhong, J. H. Zhu, L. Ding, J. X. Su, H. Yao, T. B. Wei and Y. M. Zhang, Iodine controlled pillar [5] arene-based multiresponsive supramolecular polymer for fluorescence detection of cyanide, mercury, and cysteine, *Macromolecules*, 2017, **50**, 7863–7871; (b) Y. Y. Chen, X. M. Jiang, G. F. Gong, H. Yao, Y. M. Zhang, T. B. Wei and Q. Lin, Pillararene-based AIEgens: research progress and appealing applications, *Chem. Commun.*, 2021, **57**, 284–301.
- W. J. Qu, W. T. Li, H. L. Zhang, T. B. Wei, Q. Lin, H. Yao and Y. M. Zhang, A rational designed fluorescent and colorimetric dual-channel sensor for cyanide anion based on the PET effect in aqueous medium, *Sens. Actuators, B*, 2017, **241**, 430–437.
- (a) H. H. Yang, P. P. Liu, J. P. Hu, H. Fang, Q. Lin, Y. Hong, Y. M. Zhang, W. J. Qu and T. B. Wei, A fluorescent supramolecular gel and its application in the ultrasensitive detection of CN[−] by anion– π interactions, *Soft Matter*, 2020, **16**, 9876–9881; (b) W. J. Qu, H. H. Yang, J. P. Hu, P. Qin, X. X. Zhao, Q. Lin, H. Yao, Y. M. Zhang and T. B. Wei, A novel bis-acylhydrazone supramolecular gel and its application in ultrasensitive detection of CN[−], *Dyes Pigm.*, 2021, **186**, 108949.
- S. D. Padghan, C. Y. Wang, W. C. Liu, S. S. Sun, K. M. Liu and K. Y. Chen, A naphthalene-based colorimetric and fluorometric dual-channel chemodosimeter for sensing cyanide in a wide pH range, *Dyes Pigm.*, 2020, **183**, 108724.
- R. Rajamanikandan and M. Ilanchelian, β -Cyclodextrin protected gold nanoparticle based cotton swabs as an effective candidate for specific sensing of trace levels of cyanide, *Anal. Methods*, 2019, **11**, 97–104.
- Y. M. Hijji, A. G. Elsafy, H. S. Al-Easa, B. Attili, M. Abdelrasoul, N. Mohamed and G. K. Nasrallah, Curcumin a Colorimetric and Fluorimetric Cyanide Probe in Aqueous System and Living Cells, *Anal. Methods*, 2019, **11**, 5169–5176.
- K. Rezaeian, H. Khan mohammadi and A. Talebbaigy, Detection of CN[−], Cu²⁺ and Zn²⁺ ions using a new chromone-based colorimetric chemosensor: half-adder and integrated circuits, *Anal. Methods*, 2020, **12**, 1759–1766.
- R. Kumar, V. Bhalla and M. Kumar, Cu²⁺ and CN[−] selective fluorogenic sensors based on pyrene-appended thiocalix[4]arenes, *Tetrahedron*, 2008, **64**, 8095–8101.
- M. Shahid, H. M. Chawla and P. Bhatia, A calix[4]arene based turn off/turn on molecular receptor for Cu²⁺ and CN[−] ions in aqueous medium, *Sens. Actuators, B*, 2016, **237**, 470–478.
- B. Sathish Kumar, N. N. Pati, K. V. J. Jose and P. K. Panda, Synthetic access to Calix[3]pyrroles via meso-Expansion: Hosts with diverse Guest Chemistry, *Chem. Commun.*, 2020, **56**, 5637–5640.
- P. S. Kumar, S. Ciattini, C. Laura and K. P. Elango, Fluorescent detection of Al(III) and CN[−] in solid and aqueous phases and their recognition in biological samples, *J. Mol. Liq.*, 2020, **317**, 113970.
- M. R. F. Ashworth, A Dual Colorimetric Chemosensor for Hg(II) and Cyanide ions in Aqueous Media based on Nitrobenzoxadiazole (NBD)-Antipyrine Conjugate with



- INHIBIT logic gate behavior, *Chem. Sulphonic Acids, Esters Their Deriv.*, 2006, 323–350.
- 20 S. Sharma, M. Kumari and N. Singh, C3-Symmetrical Tripodal Acylhydrazone Organogelator for the Selective Recognition of Cyanide ions in Gel and Solution phase: Practical Applications in Food Samples, *Soft Matter*, 2020, **16**, 6532–6538.
- 21 L. Chen, W. Lu, X. Wang and L. Chen, A highly selective and sensitive colorimetric sensor for iodide detection based on anti-aggregation of gold nanoparticles, *Sens. Actuators, B*, 2013, **182**, 482–488.
- 22 F. Pena-Pereira, N. Capón, I. de la Calle, I. Lavilla and C. Bendicho, Fluorescent poly(vinylpyrrolidone)-supported copper nanoclusters in miniaturized analytical systems for iodine sensing, *Sens. Actuators, B*, 2019, **299**, 126979.
- 23 M. Singh, N. Singh, J. R. Ascenso and P. M. Marcos, A Dihomooxocalix[4]arene-gold nanohybrid based colorimetric sensor for sensitive and selective detection of iodide, *Supramol. Chem.*, 2019, **31**, 313–321.
- 24 H. F. Xie, C. Wu, J. Zou, Y. X. Yang, H. Xu, Q. L. Zhang, C. Redshaw and T. Yamato, A pyrenyl-appended C3v-symmetric hexahomotrioxocalix[3]arene for selective fluorescence sensing of iodide, *Dyes Pigm.*, 2020, **178**, 108340.
- 25 P. R. Sharma, V. K. Soni, S. Pandey, G. Choudhary, A. K. Plappally and R. K. Sharma, Dipicrylhydrazine: A versatile visual anions sensor, *J. Environ. Chem. Eng.*, 2017, **5**, 2232–2239.
- 26 (a) H. M. Chawla, S. P. Singh and S. Upreti, Synthesis of calix [4]arene(amido)monocrowns and their photoresponsive derivatives, *Tetrahedron*, 2006, **62**, 9758–9768; (b) V. Saravanan, A. Kannan and P. Rajakumar, p-tert-Butylcalix[4]arene core based ferrocenyl dendrimers: Novel sensor for toxic Hg²⁺ ion even in presence of Zn²⁺, Cu²⁺ and Ag⁺ ions, *Sens. Actuators, B*, 2017, **242**, 904–911; (c) J. Xu, Y. Yang, H. Baigude and H. Zhao, New ferrocene-triazole derivatives for multi signaling detection of Cu²⁺ in aqueous medium and their antibacterial activity, *Spectrochim. Acta, Part A*, 2020, **229**, 117880; (d) M. Ramachandran, S. Anandan and M. Ashok kumar, A luminescent on-off probe based calix[4]arene linked through triazole with ruthenium(II) polypyridine complexes to sense copper(II) and sulfide ions, *New J. Chem.*, 2019, **43**, 9832–9842; (e) B. Mohan and H. K. Sharma, Synthesis of calix[6]arene and transduction of its furfural derivative as sensor for Hg(II) ions, *Inorg. Chim. Acta*, 2019, **486**, 63–68; (f) Y. Z. Chen, D. Pan, B. Chen, G. X. Wang, C. H. Tung and L. Z. Wu, Synthesis, Characterization, and Selective Sr²⁺ Sensing Study of Copper(I)-Bridged Calix[4]arene-Based Binuclear Alkynylplatinum(II) Complexes, *Eur. J. Inorg. Chem.*, 2017, **44**, 5108–5113.
- 27 (a) D. Maity, M. Bhatt, M. L. Desai, E. Suresh, M. K. Si, V. P. Boricha, B. Ganguly and P. Paul, Effect of conformation, flexibility and intramolecular interaction on ion selectivity of calix[4]arene-based anion sensors: experimental and computational studies, *Supramol. Chem.*, 2017, **29**, 600–615; (b) A. O. Ortolan, I. Østrøm, G. F. Caramori, R. L. T. Parreira, E. H. Da Silva and F. M. Bickelhaupt, Tuning heterocalixarenes to improve their anion recognition: a computational approach, *J. Phys. Chem. A*, 2018, **122**, 3328–3336; (c) F. K. W. Hau, H. S. Lo and V. W. W. Yam, Synthesis and Photophysical Studies of Calixarene-Based Alkynylplatinum(II) Terpyridine Complexes with Various Receptor Sites for Colorimetric and Luminescence Sensing of Anions, *Chem.–Eur. J.*, 2016, **22**, 3738–3749; (d) C. Parthiban, S. Ciattini, L. Chelazzi and K. P. Elango, Selective colorimetric sensing of fluoride in an aqueous solution by amino-naphthoquinone and its Co(II), Ni(II), Cu(II) and Zn(II) complexes – effect of complex formation on sensing behavior, *RSC Adv.*, 2016, **6**, 91265–91274.
- 28 H. M. Chawla, S. N. Sahu and R. Shrivastava, Synthesis and binding characteristics of novel calix [4] arene (amidocrown) diquinones, *Can. J. Chem.*, 2009, **87**, 523–531.
- 29 (a) S. B. Nimse and T. Kim, Biological applications of functionalized calixarenes, *Chem. Soc. Rev.*, 2013, **42**, 366–386; (b) C. Chen, X. Ni, H. W. Tian, Q. Liu, D. S. Guo and D. Ding, Calixarene-Based Supramolecular AIE Dots with Highly Inhibited Nonradiative Decay and Intersystem Crossing for Ultrasensitive Fluorescence Image-Guided Cancer Surgery, *Angew. Chem., Int. Ed.*, 2020, **59**, 10008–10012; (c) D. Xia, P. Wang, X. Ji, N. M. Khashab, J. L. Sessler and F. Huang, Functional Supramolecular Polymeric Networks: The Marriage of Covalent Polymers and Macrocyclic-Based Host–Guest Interactions, *Chem. Rev.*, 2020, **120**, 6070–6123.
- 30 G. Yu, K. Jie and F. Huang, Supramolecular Amphiphiles Based on Host–Guest Molecular Recognition Motifs, *Chem. Rev.*, 2015, **115**, 7240–7303.
- 31 S. Patra, D. Maity, R. Gunupuru, P. Agnihotri and P. Paul, Calixarenes: Versatile molecules as molecular sensors for ion recognition study, *J. Chem. Sci.*, 2012, **124**, 1287–1299.
- 32 C. D. Gutsche and M. Iqbal, p-tert-butylcalix[4]arene, *Org. Synth.*, 1993, **VIII**, 75–76.
- 33 C. D. Gutsche, J. A. Levine and P. K. Sujeeth, Calixarenes. 17. Functionalized Calixarenes: The Claisen Rearrangement Route, *J. Org. Chem.*, 1985, **50**, 5802–5806.
- 34 F. Unob, Z. Asfari and J. Vicens, An Anthracene-Based Fluorescent Sensor for Transition Metal Ions Derived From Calix[4]arene, *Tetrahedron Lett.*, 1998, **39**, 2951–2954.
- 35 M. Bayrakci and S. Yigiter, Synthesis of tetra-substituted calix[4]arene ionophores and the irreognition studies toward toxic arsenate anions, *Tetrahedron*, 2013, **69**, 3218–3224.
- 36 Z. Q. Shi, Y. Q. Feng, N. Song and H. W. Wang, Novel Synthesis and Characterization of N-Substituted-calix[4] azacrown Derivatives, *Synth. Commun.*, 2008, **38**, 983–990.
- 37 G. M. Sheldrick, *Saint, 5.1 edn*, Siemens Industrial Automation Inc., Madison, WI, 1995.
- 38 SADABS, *Empirical Absorption Correction Program*, University of Göttingen, Göttingen, 1997.
- 39 G. M. Sheldrick, *SHELXTL Reference Manual: Version 5.1*, Bruker AXS, Madison, WI, 1997.



- 40 G. M. Sheldrick, *SHELXL-97: Program for Crystal Structure Refinement*, University of Göttingen, Göttingen, 1997.
- 41 *Mercury 1.3, Supplied with Cambridge Structural Database*, CCDC, Cambridge, 2003.
- 42 I. D. Kuntz, F. P. Gasparro, M. D. Johnston and R. P. Taylor, Molecular Interactions and the Benesi–Hildebrand Equation, *J. Am. Chem. Soc.*, 1968, **90**, 4778–4781.
- 43 D. B. Hibbert and P. Thordarson, The death of the Job plot, transparency, open science and online tools, uncertainty estimation methods and other developments in supramolecular chemistry data analysis, *Chem. Commun.*, 2016, **52**, 12792–12805.

